

# LARGE-SCALE GENOTYPING BY NEXT GENERATION SEQUENCING: HOW TO OVERCOME THE CHALLENGES TO RELIABLY GENOTYPE INDIVIDUALS?

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## INTRODUCTION

Large-scale and reliable genotypes at individual level are required to understand the role of selective pressures on the evolution of highly polymorphic functional genes. Next-generation sequencing techniques allow for such large-scale sequencing, but post-processing treatment is still a challenge.

### OBJECTIVES

- We develop a methodology to obtain reliable genotypes after an optimized 454 sequencing and post-processing.
- We apply this methodology to an extensive dataset.
- We assess the quality of assigned genotypes through three different procedures.

### DATA

DNA extracted from hairs of 1,096 Alpine marmots (*Marmota marmota*).



### 454 SEQUENCING

Four loci of the major histocompatibility complex [1].

Well-optimized primers [1] and individual tags.

Moderate number of PCR cycles (24).

1,152 amplicons at each loci (4,608 amplicons) on 8 parts of a Roche® 454 FLX sequencing instrument.

- Low amplicon coverage to improve the cost-efficiency.

### COMPARISON OF LARGE-SCALE 454 SEQUENCING

| Species                       | cost/genotype | % assigned | Ref.              |
|-------------------------------|---------------|------------|-------------------|
| <i>Ficedula albicollis</i>    | 26.07 €       | 100%       | [2]               |
| <i>Parus major</i>            | 8.38 €        | 56%        | [3]               |
| <i>Myodes glareolus</i>       | 92.39 €       | 82%        | [4]               |
| 11 Rodent Genera              | 5.19 €        | 90%        | [5]               |
| <i>Microcebus murinus</i>     | 5.63 €        | 91%        | [6]               |
| <b><i>Marmota marmota</i></b> | <b>2.37 €</b> | <b>67%</b> | <b>This study</b> |

### 454 POST-PROCESSING PROPOSED METHODOLOGY

| Processing description                                                                           | UB reads (N amp) | UD reads (N amp) | DRB1 reads (N amp) | DRB2 reads (N amp) |
|--------------------------------------------------------------------------------------------------|------------------|------------------|--------------------|--------------------|
| Reads with individual tags and 3-bp loci markers                                                 | 152,960 (1067)   | 113,862 (1069)   | 79,114 (966)       | 113,207 (1001)     |
| Reads with > 95% and < 105% of locus sizes                                                       | 72,214 (948)     | 59,642 (944)     | 49,212 (903)       | 64,110 (901)       |
| Amplicons with > 11 reads for a given and locus                                                  | 71,611 (835)     | 58,911 (793)     | 48,291 (720)       | 63,254 (726)       |
| Reads with within a given amplicon with > 24% of allelic frequency with respect to the main read | 70,839 (835)     | 58,481 (793)     | 47,667 (720)       | 62,682 (726)       |

→ High variability of coverage per amplicon decreased the number of retained reads and amplicons.

→ The optimization of 454 post-processing increased the number of retained reads and amplicons.

→ No artefacts, PCR chimeras or new alleles were found after 454 post-processing.

### VALIDATION OF GENOTYPES

1,460 out of 3,074 obtained genotypes could be validated through intra-individual 454 sequencing repeatability (51 genotypes); 454 and Sanger sequencing repeatability (36 genotypes); and comparing the consistency of mother-father-offspring triads determined through paternity analysis (1,373 genotypes) [7].

### RESULTS

High quality MHC genotypes were obtained at the four MHC sequenced loci for 895 genotyped individuals, with a mean of  $3.43 \pm 0.96$  ( $\pm$  SD) genotypes per individual and an error rate estimated to 0.3%.

### CONCLUSIONS

- Methods applicable to any organism for which the number of loci is known.
- Our results emphasize the potential of 454 sequencing and 454 post-processing to obtain large-scale highly reliable genotypes.
- The use of the proposed methodology could improve research on the evolution of highly polymorphic functional genes.

### REFERENCES

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